

Sequencing 12S mtDNA from Neotropical Knifefishes for Genetic Barcoding

Nicole Juby¹, Nathan Lovejoy^{1,2}

1. Department of Ecology and Evolutionary Biology, University of Toronto, 2. Department of Biological Sciences, University of Toronto Scarborough



Introduction

- **Gymnotiformes** are an order of South American freshwater fishes known for their **electrosensory** systems¹
- Over 244 species, mainly in the **Amazon Basin**, with species diversity enabled by electrocommunication¹
- 5 families: Gymnotidae, Hypopomidae, Rhamphichthyidae, Apterontidae and Sternopygidae
- Organisms shed **eDNA** that can be used to assess the impacts of climate change on species abundance²
- **12S rRNA gene** (~1 kb) provides excellent taxonomic resolution for species identification³
- **Objective:** Create a reference library of Gymnotiformes 12S sequences for use in genetic barcoding and eDNA studies



Figure 1. Electric eel (*Electrophorus electricus*). Image: Steven G. Johnson

Methods

PCRs were run to amplify the 12S gene from **150 species** using the following primers⁴:

Forward	5'-CAA AGG CTT GGT CCT GAC-3'
Reverse	5'-AGC ATT CCC TTG CGG TAC-3'

Table 1. Forward and reverse primers used for 12S PCR

Results were verified via gel electrophoresis for **30 minutes at 90V**, ensuring that a fragment of ~**1000 bp** had been amplified

If no band appeared for a particular sample, another sample from the same species was used to reattempt amplification

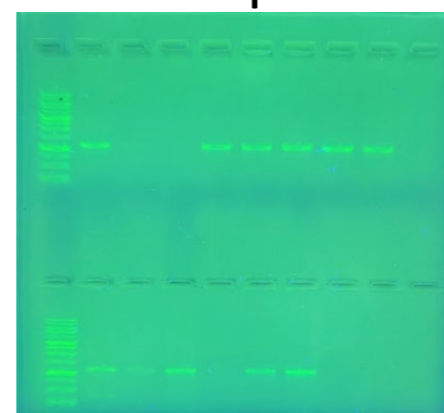


Figure 2. Illuminated gel showing 10 well-amplified 12S samples

Successfully amplified fragments were sent for sequencing at The Centre for Applied Genomics (TCAG)

Returned chromatograms were assembled and trimmed to form a **consensus sequence** for each species in Geneious Prime⁵

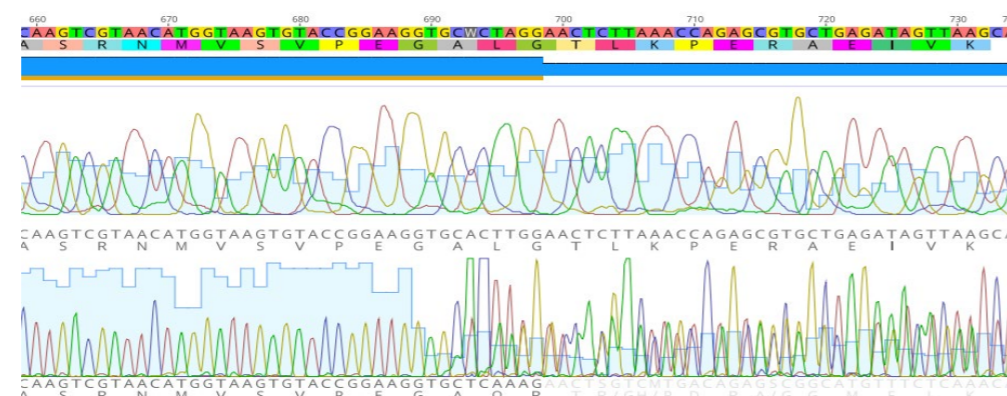


Figure 3. Portion of sequence assembly for *Gymnotus obscurus* (bp 650-735)

Results and Future Directions

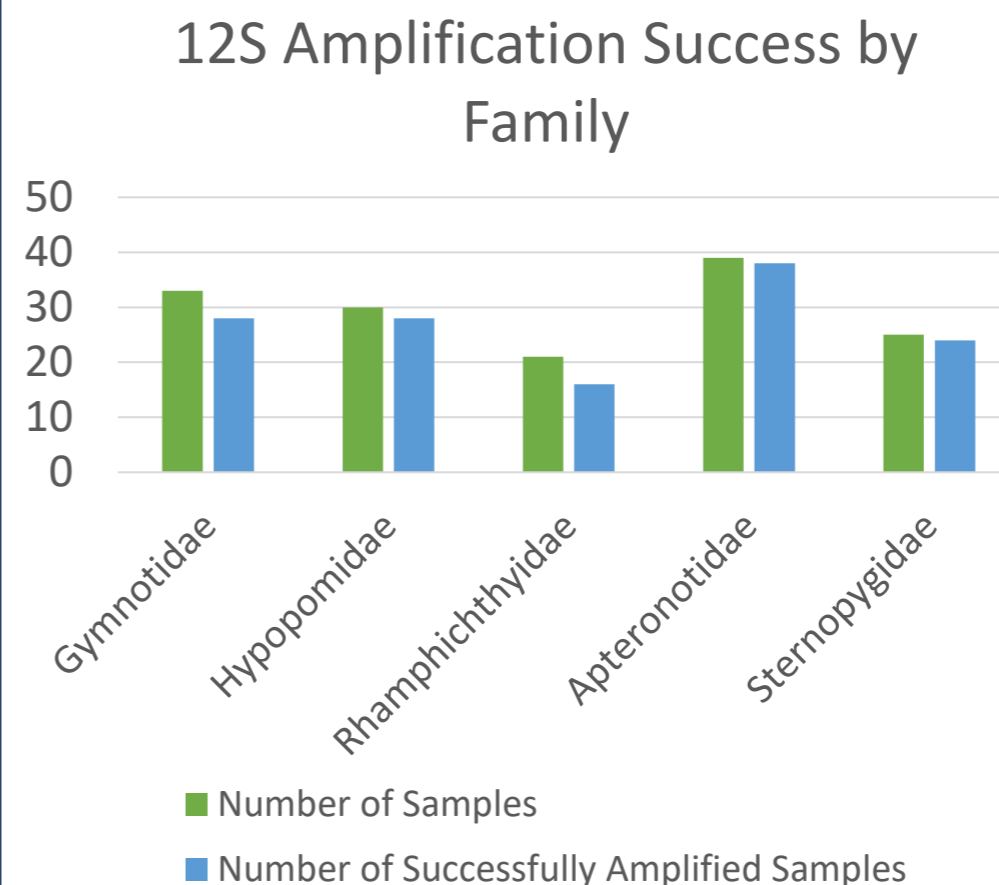


Figure 4. PCR success by family

- **103 sequences** obtained
- Average sequence length: **1001 bp**
- Family with lowest success rate (79.19%) was **Rhamphichthyidae**, which can be referenced to develop new primers that better target 12S in currently unamplified species

References and Acknowledgements

1. Crampton WGR. 2019. Electroreception, electrogenesis and electric signal evolution. *J Fish Biol.* 95(1): 92-134
2. Thomsen PF, Willerslev E. 2015. Environmental DNA – An emerging tool in conservation for monitoring past and present biodiversity. *Biol Conserv.* 183: 4-18
3. Milan DT, Mendes IS, Damasceno JS, Teixeira DF, Sales NG, Carvalho DC. 2020. New 12S metabarcoding primers for enhanced Neotropical freshwater fish biodiversity assessment. *Sci Rep-UK.* 10: 17966
4. Van Nynatten A, Gallage KS, Lujan NK, Mandrak NE, Lovejoy NR. 2023. Ichthyoplankton metabarcoding: An efficient tool for early detection of invasive species establishment. *Mol Ecol Resour.* 23(6): 1319-1333
5. Geneious Prime 2022.1.1 (<https://geneious.com>)

I would like to thank the Centre for Global Change Science for facilitating this research opportunity, in addition to the members of the Lovejoy Lab for their help and support. I would also like to thank Kaelyn Gomes for her assistance in running PCRs, and finally Dr. Nathan Lovejoy for providing me with the opportunity to participate in this project.